THE CHEMISTRY OF THE LEUCOMYCINS. II.

STRUCTURE AND STEREOCHEMISTRY OF LEUCOMYCIN A3.

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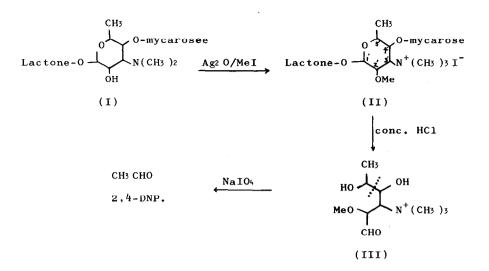
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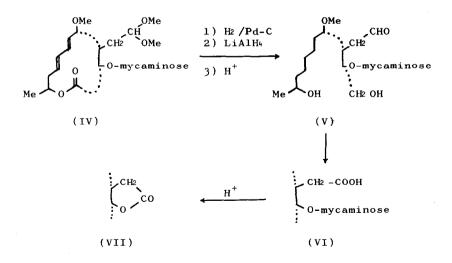
In the previous communication¹⁾, it was described that leucomycin A3 is a macrolide containing mycaminose and isovaleryl mycarose. In this paper we propose the full structure of leucomycin A3 (I) and also the stereochemistry of the mycarosidic linkages of spiramycin and magnamycin²⁾ in relation to leucomycin A3. Dehydroleucomycin A3¹⁾ obtained from <u>I</u> by oxidation with MnOz is identical with magnamycin B³⁾ as compared by IR, UV and NMR spectra, and behavior on thin layer chromatography. From these experiments, the structure of <u>I</u> becomes clear. Further support for the structure of <u>I</u> follows from the following experiments.

The pka' values of <u>I</u>, diacetyl leucomycin A3, demycarosyl leucomycin A3, and triacetyl demycarosyl leucomycin A3, 6.70, 5.69, 7.80 and 5.35, respectively¹⁾, suggest that mycarose is attached to either C-2' or C-4' of the mycaminose, i.e., a position neighbouring the dimethylamino group⁴⁾. The position of the mycarose residue can be fixed at C-4' since treatment of 0,0-dimethyl leucomycin A3 methyliodide (II) with conc. HCl affords <u>III</u>, which when cleaved by sodium periodate provides acetaldehyde in good yield.

Rotations were measured in chloroform solution at 25°C, UV spectra were taken in methanol solution and pka' values were measured in 50% ethanol. Satisfactory analyses were obtained for all compounds for which molecular formulae are given.



The position of -CH2 CHO group of \underline{I}^{1} is confirmed at the neighboring position of the mycaminoside linkage by the method of Woodward⁴⁾. As shown in the scheme, methyl demycarosyl leucomycin A3 dimethylacetal (IV) is reduced by catalytic hydrogenation to give tetrahydro methyl demycarosyl leucomycin A3¹⁾, which is further reduced with LiAlH4 to yield a triol (V). This aldehyde can be oxidized with H2 O2 to afford an acid (VI), which upon heating with dil. HCl yields a \underline{X} -lactone (VII) showing an absorption at 1770 cm⁻¹ in the IR spectrum. These experiments enable one to confirm the position of he aldehyde group.



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Hexahydro leucomycin A3 (VIII), C42 H75015N, $(\alpha)_{D}$ -48.8° (c 1.35) is obtained by catalytic hydrogenation of \underline{I} , and shows no aldehyde group in the IR and NMR spectra. Treatment of VIII with HCl in methanol yields methyl 4-0-isovaleryl mycaroside (IX) and demycarosyl hexahydro leucomycin A3 (X), C30H55O11N, $[\alpha]_n$ -10.4° (c 1.33), and when the latter is warmed with 10 N. NaOH the acid (XI) is obtained as a colorless liquid, $\lambda_{max}^{0.01}$ N NaOH 265 mµ ($E_{1cm}^{1\%}$ 650), γ_{max} 1700, 1630 and 1605 cm⁻¹. The same alkaline treatment of hexahydroforocidine (obtained from spiramycin) gives an acid having the same UV spectrum, thus suggesting the presence of an $\alpha, \beta, \gamma, \delta$ -unsaturated \underline{I} -methoxy acid system 4)5 in <u>XI</u>. On the other hand, when decahydro leucomycin A3 (XII), C35H69O13N, $(\alpha)_{D}$ -55.5° (c 1.33), obtained from tetrahydro leucomycin A3 by reduction with LiAlH4, is submitted to the same HC1/ MeOH treatment as described above for VIII, the demycarosyl derivative XIII, C28H57O10N, $(\alpha)_n$ -15.0° (c 1.33), is obtained. Treatment of <u>XIII</u> with 10 N. NaOH fails to give an acid corresponding to the compound (XI), as evidenced by the UV spectrum. From these experiments, it is evident that the carbonyl group of the original lactone is converted to the carboxylic group of the acid (XI).

In the NMR spectrum of \underline{I} (Fig. 1), the signal at 4.05 ppm., which is shifted to a lower field upon acetylation, can be assigned to the C9 proton. The C9 proton is coupled to the C10 proton at 5.60 ppm. (J 8.9 cps) and the

$\frac{\text{Hz}/\text{PtOz}}{\underline{I}} \text{Hexahydro leucomycin A3}$	HC1-MeOH ────→	Methyl 4-0-isovaleryl
(VIII)		mycaroside (IX)
H2 /Pd-C		+
Tetrahydro leucomycin A3		Demycarosyl hexahydro
LIA1H4		leucomycin A3 (X)
Decahydro leucomycin A3 (XII)		10 N. NaOH
HC1-MeOH		10 N. NaOH
Demycarosyl decahydro leucomycin A3	(XIII)	-CH=C-CH=CH-COOH
		OMe
		(XI)

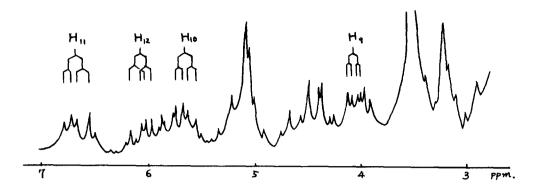
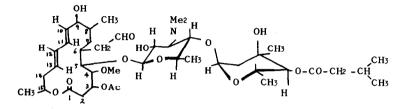


Fig. 1. NMR spectrum of leucomycin A3 (CDC13; 100 Mc)

C8 proton (J 4.2 cps). Although the signal of the C8 proton cannot be detected, C8 should be a tertiary carbon from the quartet nature of the C9 proton peak. Signals of the C11 proton at 6.60 ppm. (1 H, J11,10 15.4 cps, J11,12 10.0 cps) and the C12 proton at 6.05 ppm. (J12,11 10.0 cps, J12,13 15.2 cps) in Fig. 1 indicate a <u>trans-trans</u> configuration for the diene system⁶.

Finally, the stereochemistry of the glycoside linkage of leucomycin A3 will be discussed. In the NMR spectra of \underline{I} and related compounds, a 1 H doublet at 4.30 ppm. may be attributed to the anomer proton (1' H) (J 7.4 cps), and this suggests a β -configuration of the mycaminosidic linkage (Fig. 1). On the other hand, the stereochemistry of the mycaminoside linkage cannot be determied in a straight forward from the NMR, because many signals appear around 5.00 ppm. On the other hand, since diacetyl leucomycin $A_3^{(1)}$ has only one tertiary hydroxyl group, a comparison of the IR spectra in **CG1**4 of the acetate and α - and β -methyl 4-0-isovaleryl mycaroside (IXa and IXb)¹⁾ makes it possible to examine the formation of intramolecular hydrogen bonding. It is found that the $\underline{\beta}$ -isomer has an absorption peak at 3615 cm⁻¹ due to a free hydroxyl group. On the contrary the α -isomer has an absorption peak at 3530 cm^{-1} which is due to a hydroxyl group hydrogen bonded to the glycosidic methoxyl oxygen. The hydroxyl group absorptions of acetyl leucomycin A3, acetyl spiramycin and acetyl magnamycin appear at 3515, 3515, and 3518 $\rm cm^{-1}$, respectively. It is therefore concluded that the mycarosidic

linkages of leucomycin A3, spiramycin and magnamycin adopt the $\underline{\alpha}$ -forms. On the basis of these data, the structure shown below is proposed for leucomycin A3.





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REFERENCES

 S. Ōmura, H. Ogura and T. Hata, Tetrahedron letters, submitted.
After submission of this communication, a paper by W. D. Celmer J. Am. Chem. Soc., <u>88</u>, 5028 (1966) has been published in which the same stereochemistry for the sugar constituents has been derived.
F. A. Hochstein and K. Murai, J. Am. Chem. Soc., <u>76</u>, 5080 (1954).
R. B. Woodward, Angew. Chem., <u>69</u>, 50 (1957).
R. Paul, S. Tchelitcheff, Bull. soc. chim. France, 150 (1960).
L. M. Jackman "Application of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry" Pergamon Press, New York, N. Y. 1959, p. 85.